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Bioorganic & Medicinal Chemistry 14 (2006) 7359-7361

Bioorganic & Medicinal Chemistry

#### Addendum and Corrigendum

# Addendum and corrigendum to "Inhibition of *O*-GlcNAcase by PUGNAc is dependent upon the oxime stereochemistry"

[Bioorg. Med. Chem. 14 (2006) 837–846]

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Available online 30 August 2006

### 1. Confirmation of PUGNAc oxime stereochemistry and *O*-GlcNAcase inhibition

Based on the previous work of Vasella and co-workers, the assignment of **3a** as Z-PUGNAc is confirmed via alignment of NMR signals supporting our conclusion that Z-PUGNAc is the active inhibitor of O-GlcNAcase (included are both proton and carbon spectroscopic data for compounds **6a** and **3a** with full assignments based upon COSY, HMQC, and HMBC experiments). Further, a recent crystallographic analysis of PUGNAc within the active site of a related hexosaminidase (*CpNagJ*) confirms this conclusion by showing PUGNAc as the *Z* isomer in a binding position that would not accommodate the *E* stereochemistry. However, recent results of ongoing studies of the presumptive PUGNAc stereoisomers prompt us to revise two of our conclusions.

#### 2. Regarding the assignment of Z-PUGNAc (3a)

While the stability profiles for Z-PUGNAc and E-PUGNAc are conveniently explained by the Beckmann rearrangement, the observed mass analysis and proton NMR data for the products of the presumed Beckmann rearrangement are *more* consistent with the products that would arise from simple hydrolysis of **3a**. Full characterisation of such a hydrolysis product has *not* been achieved, and thus the exact nature of the mechanism by which **3a** breaks down should be considered tentative. However, the realisation that Z-PUGNAc likely breaks down via

DOI of original article: 10.1016/j.bmc.2005.09.013.

hydrolysis led to us question the apparent resistance of **3b** to a similar hydrolysis event and eventually led to us question the structure of **3b**.

#### 3. Regarding the structural assignment of compound 3b

The spectroscopic data for 3b potentially match the predicted values for E-PUGNAc; however, a more accurate match for the proton and carbon spectrum is the related 1,4-lactone species (a possible by-product of the synthesis of PUGNAc as previously stated by Vasella and coworkers<sup>1</sup>). A COSY correlation spectrum of **3b** in very dry DMSO-d<sub>6</sub> provides for the coupling between the hydroxyl protons and the sugar methine and methylene protons. This spectrum clearly shows the C5 methine proton coupled to a hydroxyl proton; no associated coupling between the C4 methine proton and any hydroxyl signal was noted. A HMBC correlation spectrum of 6b subsequently confirmed the presence of the 1.4-lactone triacetate product via the three bond coupling of the C5 methine proton and the carbonyl carbon of an acetate; no associated coupling between an acetate carbonyl and the C4 methine proton was noted (the aforementioned COSY spectrum of 3b and HMBC spectrum of 6b are included here as Figs. 1 and 2). Thus, there is clear evidence for the existence of the 1,4-lactone product rather than E-PUGNAc, and compounds 6b and 3b should be listed as shown below (included are both proton and carbon spectroscopic data for compounds 6b and 3b with full assignments based upon COSY, HMQC, and HMBC experiments). Therefore, while the conclusion that Z-PUGNAc is the primary inhibitor remains valid, the activity assigned to compound 3b should be associated with the 1,4-lactone product shown below and not E-PUGNAc.

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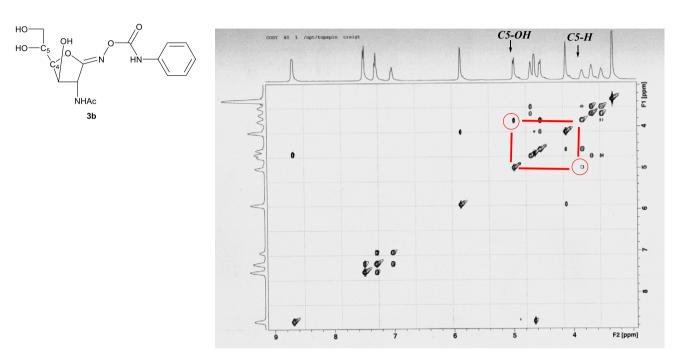


Figure 1. COSY spectrum of 3b.

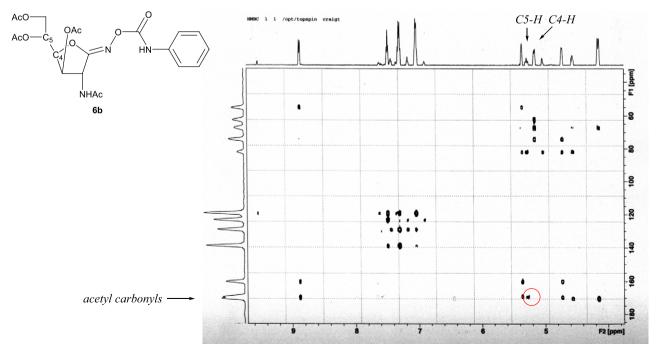


Figure 2. HMBC spectrum of 6b.

## 4. *O*-(2-Acetamido-3,4,5-tri-*O*-acetyl-D-glucopyranosylidene)amino-*Z*-*N*-phenylcarbamoyl (6a)

<sup>1</sup>H NMR (DMSO- $d_6$ ) δ 1.87 (s, 3H, Ac-CH<sub>3</sub>), 2.00 (s, 3H, Ac-CH<sub>3</sub>), 2.03 (s, 3H, Ac-CH<sub>3</sub>), 2.05 (s, 3H, Ac-CH<sub>3</sub>), 4.20 (dd,  $J_{\text{HH}} = 12.9$ ,  $J_{\text{HH}} = 2.4$  Hz, 1H, C6-H), 4.38 (dd,  $J_{\text{HH}} = 12.9$ ,  $J_{\text{HH}} = 4.4$  Hz, 1H, C6-H), 4.66 (ddd,  $J_{\text{HH}} = 9.4$ ,  $J_{\text{HH}} = 4.3$  Hz,  $J_{\text{HH}} = 2.4$ , 1H, C5-H), 4.79 (dd appears as t,  $J_{\text{HH}} = 7.9$ ,  $J_{\text{HH}} = 7.9$  Hz, 1H, C2-H), 5.24 (dd,  $J_{\text{HH}} = 9.4$ ,  $J_{\text{HH}} = 8.0$  Hz, 1H, C4-H), 5.34 (dd appears as t,  $J_{\text{HH}} = 8.0$ ,  $J_{\text{HH}} = 8.0$  Hz, 1H, C3-H), 7.02–7.05 (m, 1H, Ar-H), 7.29–7.32 (m, 2H, Ar-H), 7.48–7.49 (m, 2H, Ar-H), 8.61 (d,  $J_{\text{HH}} = 7.68$  Hz, 1H, NHAc), 9.74 (s, 1H, PhNH); <sup>13</sup>C NMR (DMSO- $d_6$ ) δ 18.8 (COCH<sub>3</sub>), 18.1 (COCH<sub>3</sub>), 18.9 (COCH<sub>3</sub>), 20.9 (COCH<sub>3</sub>), 47.1 (C2), 60.0 (C6), 65.5 (C4), 69.3 (C3), 74.2 (C5), 117.3, 121.4, 127.2, 136.8, 149.8 (OCONH), 154.1 (C1), 167.5 (COCH<sub>3</sub>), 167.7 (COCH<sub>3</sub>), 168.0 (COCH<sub>3</sub>), 168.4 (COCH<sub>3</sub>).

## 5. (*Z*)-1-(4-Acetamido-3,5,6-triacetyl-5-(hydroxyimino)tetrahydrofuran-2-yl)ethane-1,2-diyl (6b)

<sup>1</sup>H NMR (DMSO- $d_6$ ) δ 1.90 (s, 3H, Ac-CH<sub>3</sub>), 1.99 (s, 3H, Ac-CH<sub>3</sub>), 2.04 (s, 3H, Ac-CH<sub>3</sub>), 2.05 (s, 3H, Ac-CH<sub>3</sub>), 4.13 (dd,  $J_{\text{HH}}$  = 12.4,  $J_{\text{HH}}$  = 5.8 Hz, 1H, C6-H), 4.55 (dd,  $J_{\text{HH}}$  = 12.4,  $J_{\text{HH}}$  = 2.6 Hz, 1H, C6-H), 4.72 (dd,  $J_{\text{HH}}$  = 7.6,  $J_{\text{HH}}$  = 1.7 Hz, 1H, C2-H), 5.15 (dd,  $J_{\text{HH}}$  = 8.2,  $J_{\text{HH}}$  = 4.3 Hz, 1H, C4-H), 5.26 (ddd,  $J_{\text{HH}}$  = 8.3,  $J_{\text{HH}}$  = 5.6 Hz,  $J_{\text{HH}}$  = 2.6, 1H, C5-H), 5.34 (dd,  $J_{\text{HH}}$  = 4.3,  $J_{\text{HH}}$  = 1.8 Hz, 1H, C3-H), 7.02–7.04 (m, 1H, Ar-H), 7.28–7.31 (m, 2H, Ar-H), 7.47–7.48 (m, 2H, Ar-H), 8.90 (d,  $J_{\text{HH}}$  = 7.56 Hz, 1H, NHAc), 9.84 (s, 1H, PhNH); <sup>13</sup>C NMR (DMSO- $d_6$ ) δ 18.5 (COCH<sub>3</sub>), 18.9 (COCH<sub>3</sub>), 19.0 (COCH<sub>3</sub>), 20.7 (COCH<sub>3</sub>), 53.1 (C2), 60.4 (C6), 65.3 (C5), 72.2 (C3), 80.0 (C4), 117.6, 121.4, 127.2, 136.9, 149.8 (OCONH), 158.3 (C1), 167.3 (COCH<sub>3</sub>), 167.5 (COCH<sub>3</sub>), 168.6 (COCH<sub>3</sub>).

## 6. *O*-(2-Acetamido-2-deoxy-*D*-glucopyranosylidene)-amino-*Z*-*N*-phenylcarbamoyl (3a)

<sup>1</sup>H NMR (DMSO- $d_6$ ); δ 1.89 (s, 3H, Ac-CH<sub>3</sub>), 3.58 (dd,  $J_{\rm HH}$  = 8.5,  $J_{\rm HH}$  = 7.8 Hz, 1H, C4-H), 3.64 (dd,

 $J_{\rm HH}$  = 8.2,  $J_{\rm HH}$  = 8.0 Hz, 1H, C3-H), 3.65–3.70 (m, 1H, C6-H), 3.75–3.77 (m, 1H, C6-H), 3.92–3.95 (m, 1H, C5-H), 4.37 (dd appears as t,  $J_{\rm HH}$  = 8.3,  $J_{\rm HH}$  = 8.3 Hz, 1H, C2-H), 4.93 (br s, 1H, C6-OH), 5.55 (br s, 2H, C3-OH and C4-OH), 7.00–7.02 (m, 1H, Ar-H), 7.27–7.38 (m, 2H, Ar-H) 7.48–7.49 (m, 2H, Ar-H), 8.34 (d,  $J_{\rm HH}$  = 8.1 Hz, 1H, NHAc), 9.62 (s, 1H, PhNH); <sup>13</sup>C NMR (D<sub>2</sub>O at 15 °C);  $\delta$  23.8 (COCH<sub>3</sub>), 51.8 (C2), 60.7 (C6), 69.2 (C4), 72.9 (C3), 83.1 (C5), 119.3, 123.5, 129.4, 139.2, 152.4 (OCONH), 158.8 (C1), 169.9 (COCH<sub>3</sub>).

## 7. (*Z*)-*N*-(5-(1,2-Dihydroxyethyl)-4-hydroxy-2-(hydroxyimino)tetrahydrofuran-3-yl)acetamide (3b)

<sup>1</sup>H NMR (DMSO- $d_6$ ); δ 1.87 (s, 3H, Ac-CH<sub>3</sub>), 3.50–3.53 (m, 1H, C6-H), 3.60–3.61 (m, 1H, C6-H), 3.82–3.85 (m, 1H, C5-H), 4.11–4.12 (m, 1H, C3-H), 4.54 (dd,  $J_{\rm HH}$  = 8.2,  $J_{\rm HH}$  = 3.1 Hz, 1H, C4-H), 4.63 (d,  $J_{\rm HH}$  = 7.7, 1H, C2-H), 4.69–4.71 (m, 1H, C6-OH), 4.97–4.99 (m, 1H, C5-OH), 5.82 (br s, 1H, C3-OH), 7.00–7.03 (m, 1H, Ar-H), 7.27–7.30 (m, 2H, Ar-H), 7.48–7.49 (m, 2H, Ar-H), 8.66 (d, J = 7.7 Hz, 1H, NHAc), 9.73 (br s, 1H, PhNH); <sup>13</sup>C NMR (DMSO- $d_6$ ); δ 23.1 (COCH<sub>3</sub>), 57.4 (C2), 63.6 (C6), 68.8 (C5), 73.3 (C3), 86.0 (C4), 119.4, 123.5, 129.4, 139.2, 152.3 (OCONH), 162.6 (C1), 169.5 (COCH<sub>3</sub>).

#### Acknowledgment

The authors express gratitude to Dr. Spencer Knapp for elucidating the likelihood of the hydrolysis of **3a** and for helpful discussions during subsequent correction of the structures of **6b** and **3b**.

#### References and notes

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